

# Some comments on dose, signal to noise, damage, and heating

Chris Jacobsen

Argonne/Northwestern

DLSR workshop, Dec. 10, 2013

# Signal to noise and required number of photons

- Simple photon statistics with known contrast:

$$\text{SNR} = \frac{\text{Signal}}{\text{Noise}} = \frac{\bar{n}|I_f - I_b|}{\sqrt{(\sqrt{\bar{n}I_f})^2 + (\sqrt{\bar{n}I_b})^2}} = \sqrt{\bar{n}} \frac{|I_f - I_b|}{\sqrt{I_f + I_b}} = \sqrt{\bar{n}}\Theta$$

where  $\Theta$ =contrast parameter,  $I_f$ =intensity of feature,  $I_b$ =intensity of background.

- Thus required number of incident photons  $\bar{n}$  is

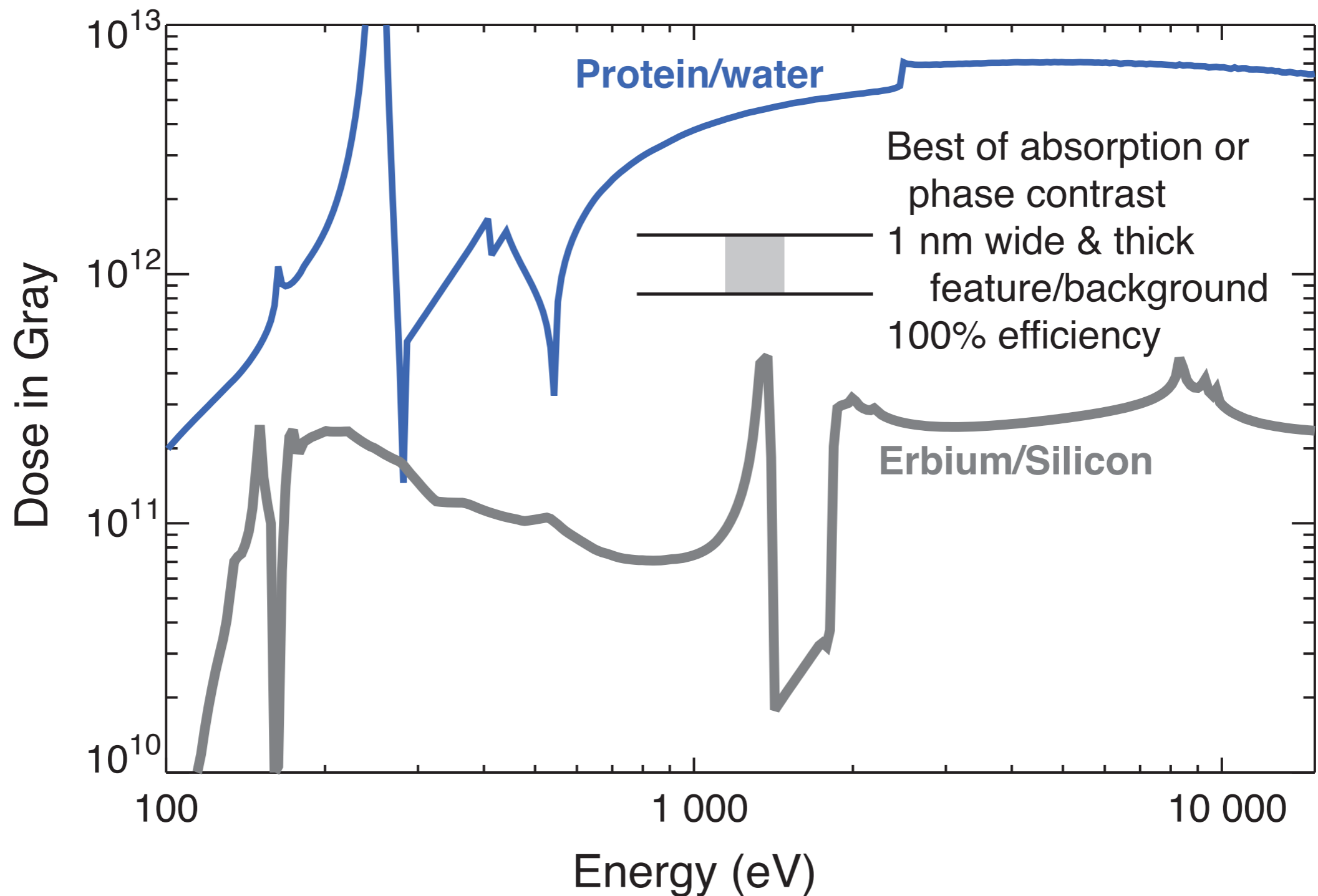
$$\bar{n} = \frac{\text{SNR}^2}{\Theta^2}$$

- Example: 10 nm protein in ice at 520 eV via absorption contrast
  - Protein has linear absorption coefficient (LAC) of 1/9.900  $\mu\text{m}$ , so 10 nm has  $I_f = \exp[-0.010/9.900]=0.99899$
  - Ice has LAC of 0.717  $\mu\text{m}$ , so 10 nm has  $I_b = \exp[-0.010/0.717]=0.98615$
  - Contrast parameter is  $\Theta = (.99899 - .98615) / (.99899 + .98615)^{1/2} = .00911$
  - So with SNR=5 one requires  $\bar{n} = (5)^2 / (.00911)^2 = 3 \times 10^5$  incident photons
- See e.g., Sayre *et al.*, *Ultramicroscopy* **2**, 337 (1977); Sayre *et al.*, *Science* **196**, 1339 (1977)



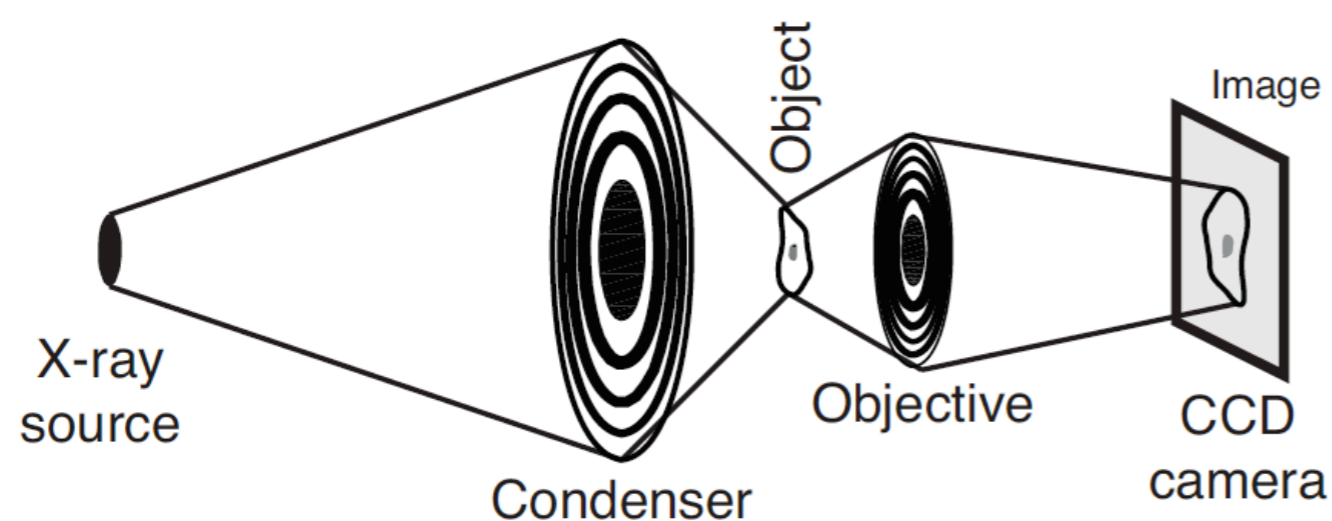
# Estimates of dose for 1 nm resolution imaging

Absorbed dose correlates well with damage across a wide spectrum of ionizing radiation. Example: VUV, e-beam, x-ray exposure of photoresists.

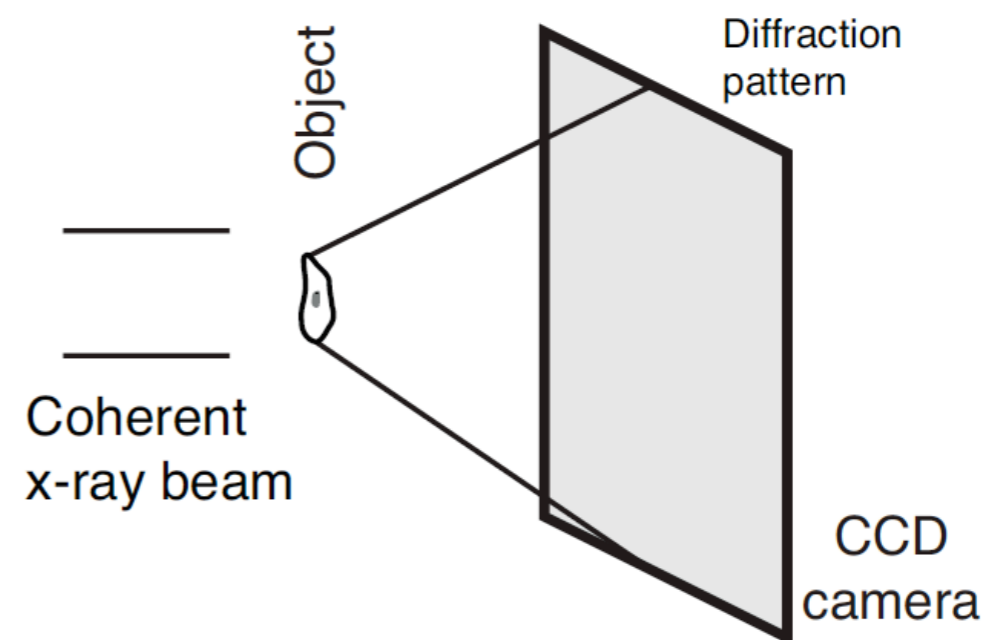


# Comparison: TXM versus XDM

- TXM: assume zone plate with 20 nm outermost zone width (mean MTF  $\sim 20\%$ ), 10% efficiency. Net throughput:  $\sim 2\%$ .
- XDM: assume 100% efficient detector.



TXM: transmission x-ray microscope



XDM: x-ray diffraction microscope

X. Huang *et al.*, *Optics Express* **17**, 13541 (2009)

# SNR from unknown objects

- What if we don't know  $I_f$  and  $I_b$ ?
- Let's say we have two noisy images  $I_1$  and  $I_2$ , but we know they are of the same object.
- Calculate correlation  $r$ :

$$r = \frac{\langle (I_1 - \langle I_1 \rangle)(I_2 - \langle I_2 \rangle)^* \rangle}{\sqrt{\langle (I_1 - \langle I_1 \rangle)^2 \rangle \langle (I_2 - \langle I_2 \rangle)^2 \rangle}}$$

- We find  $\text{SNR} = \sqrt{r/(1-r)}$
- Note: square root of expression of Frank and Al-Ali, *Nature* **256**, 376 (1975)

## Two simulated “cells” at 540 eV

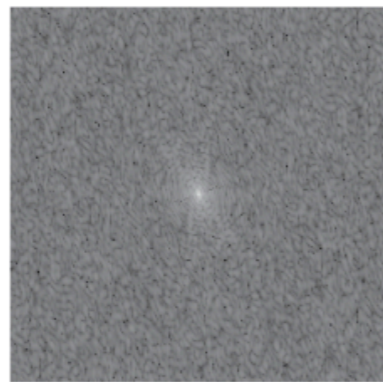
- Cell A: 0-500 nm random protein thickness
- Cell B: protein spheres, bars in ice (3D exit wave)

Exit wave

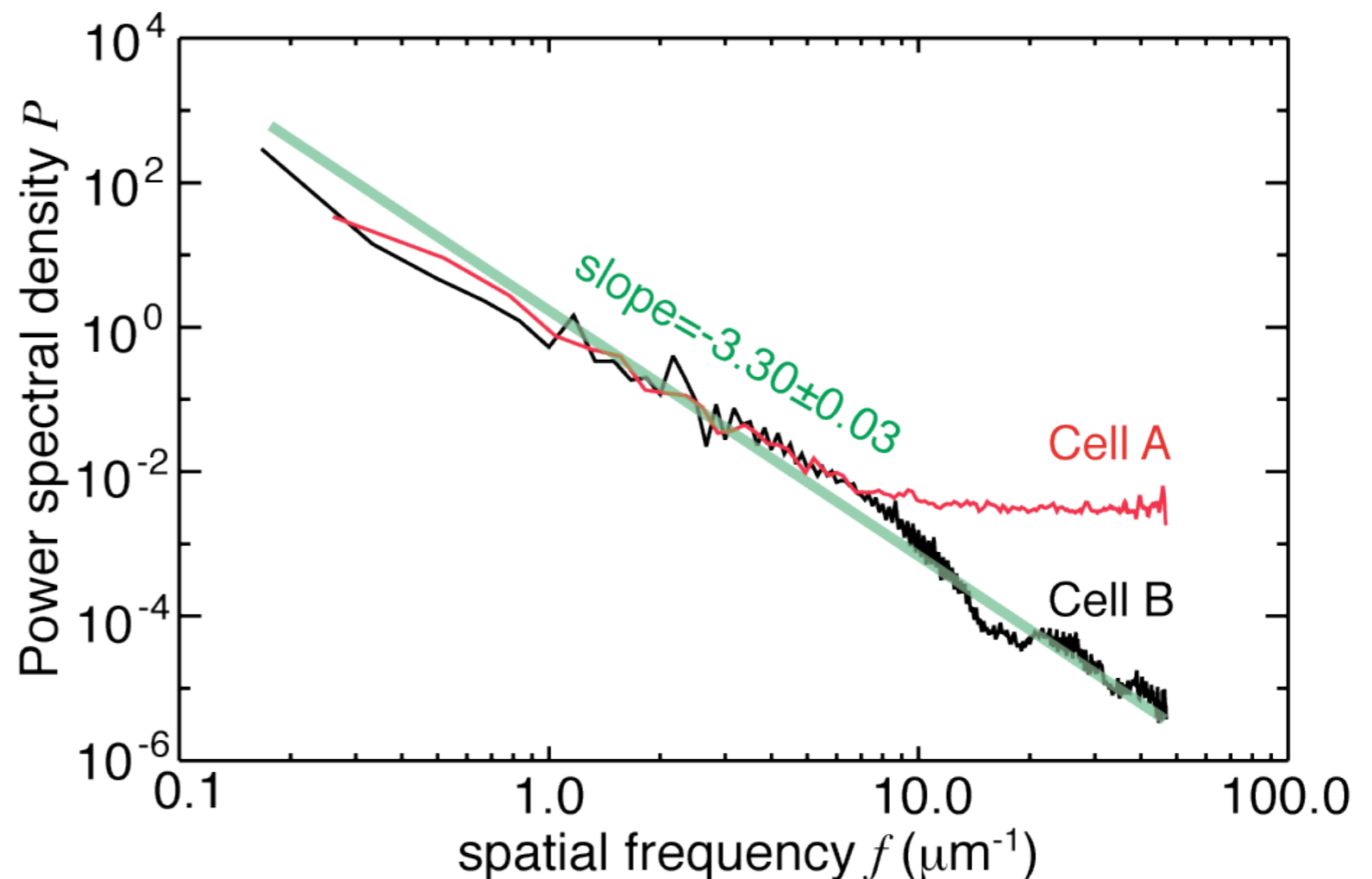
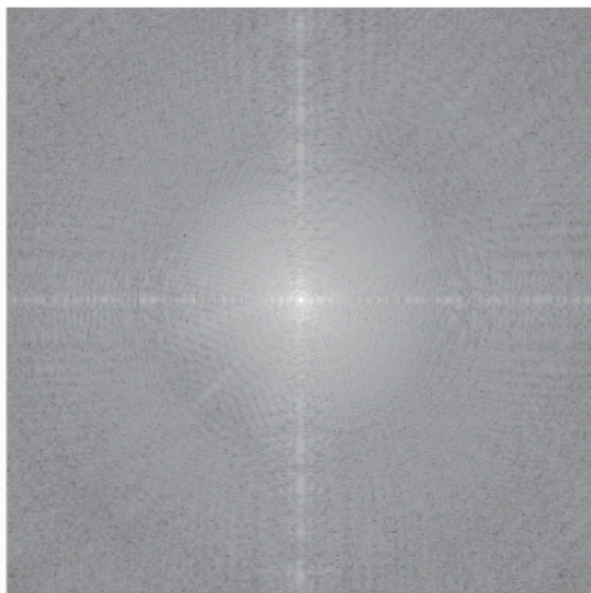
Diffraction pattern



Cell A



Cell B

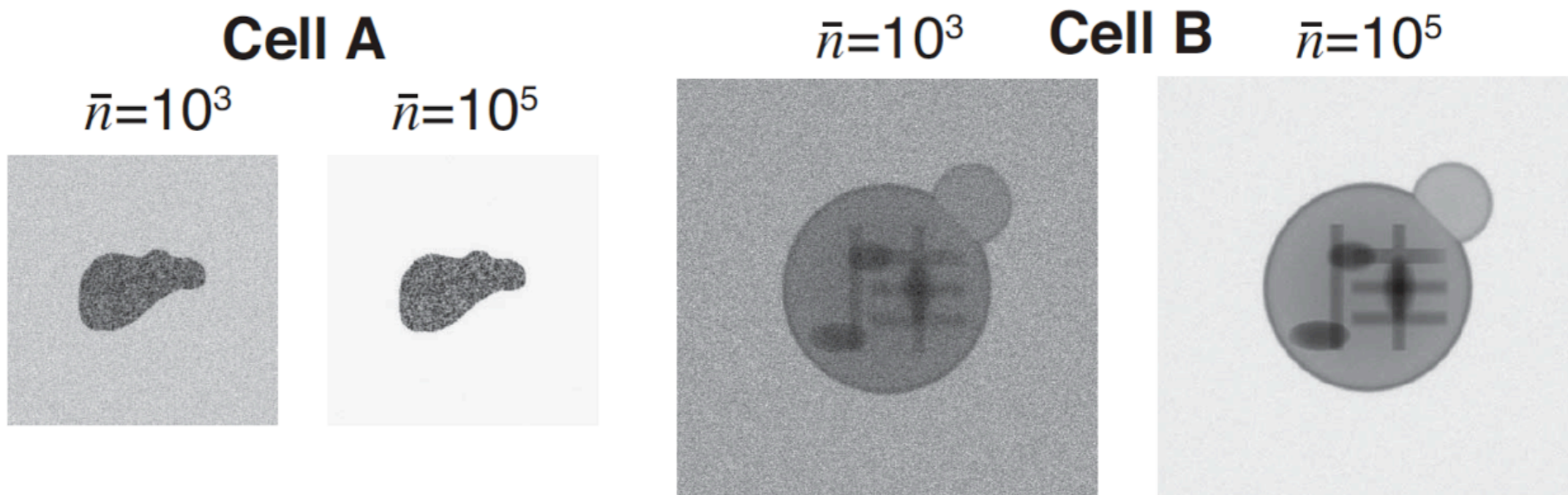


X. Huang *et al.*, *Optics Express* **17**, 13541 (2009)

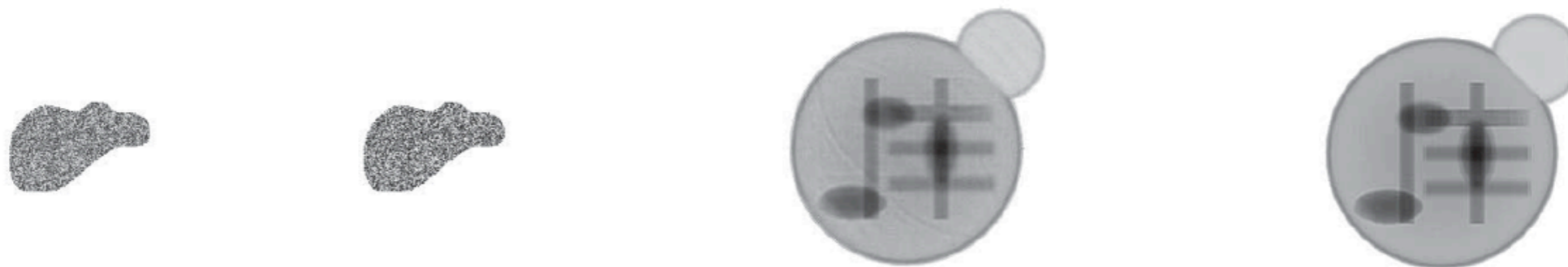
# Fake “cells” via TXM and XDM

- XDM: assume perfect support

Zone plate  
imaging  
(TXM)



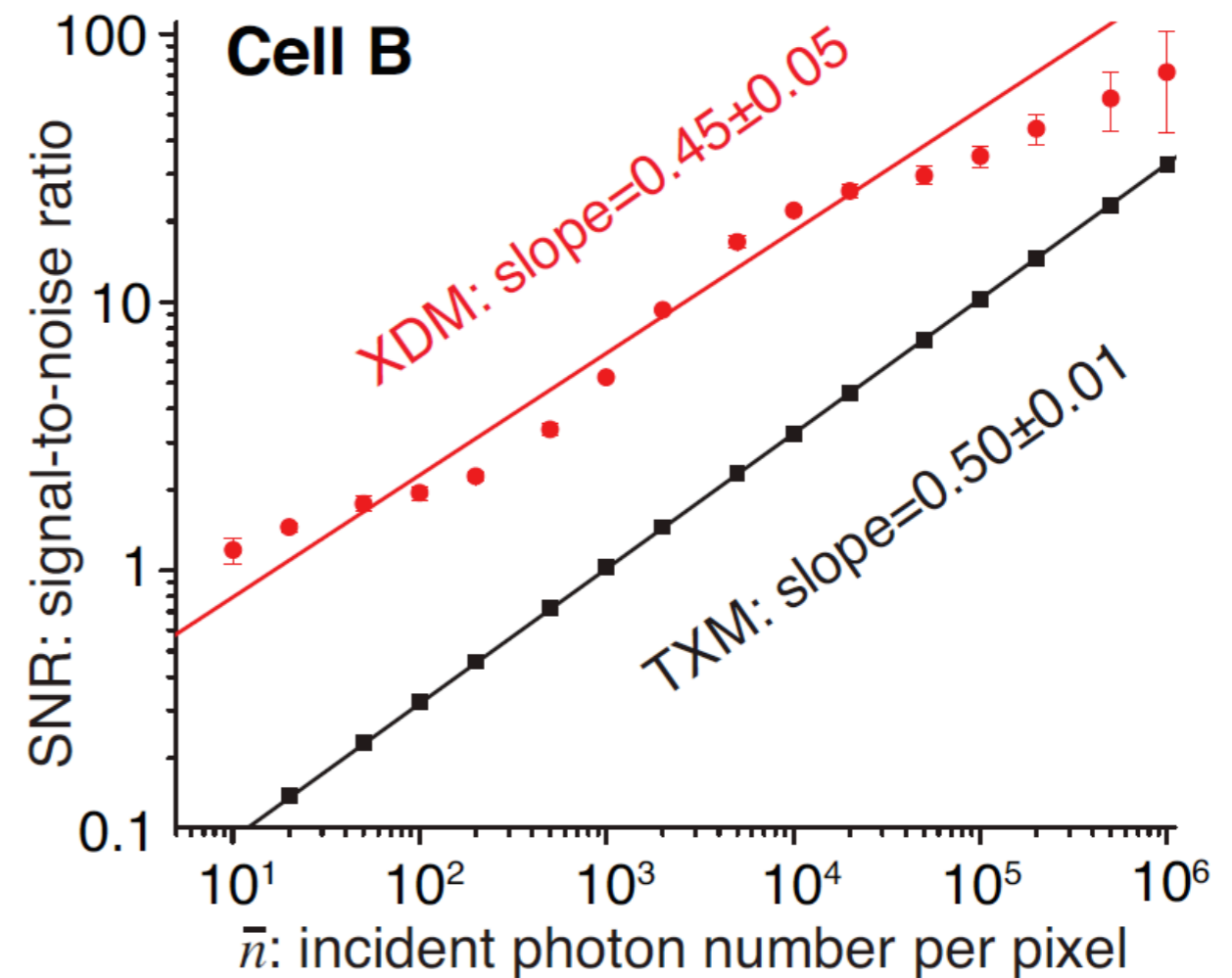
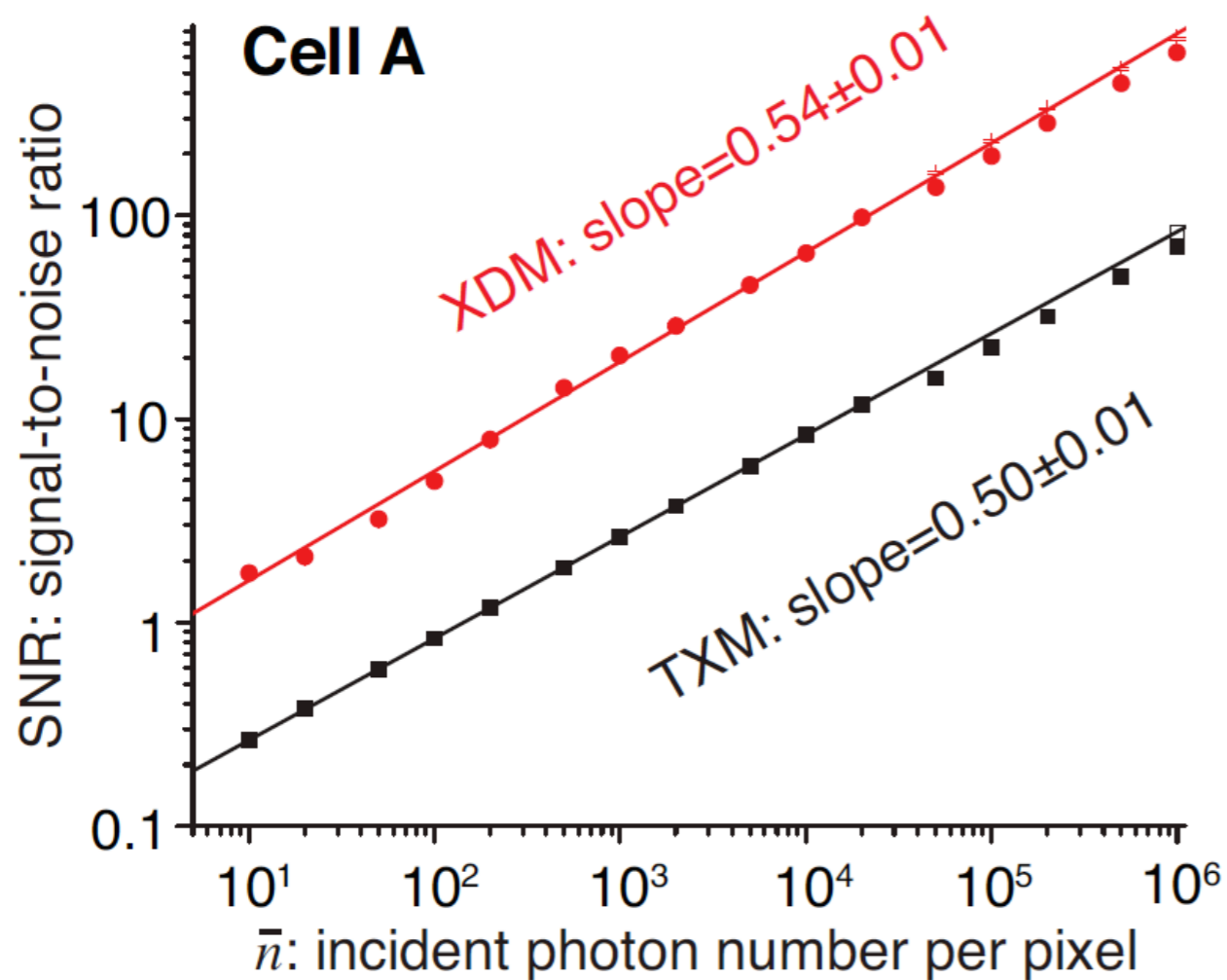
Diffraction  
microscopy  
(XDM)



X. Huang *et al.*, *Optics Express* **17**, 13541 (2009)

# SNR versus exposure: results

- TXM: net throughput  $\sim 2\%$ , or  $1/50$ . Expect SNR to be  $\sim \sqrt{50}$  or  $\sim 7$  times lower.

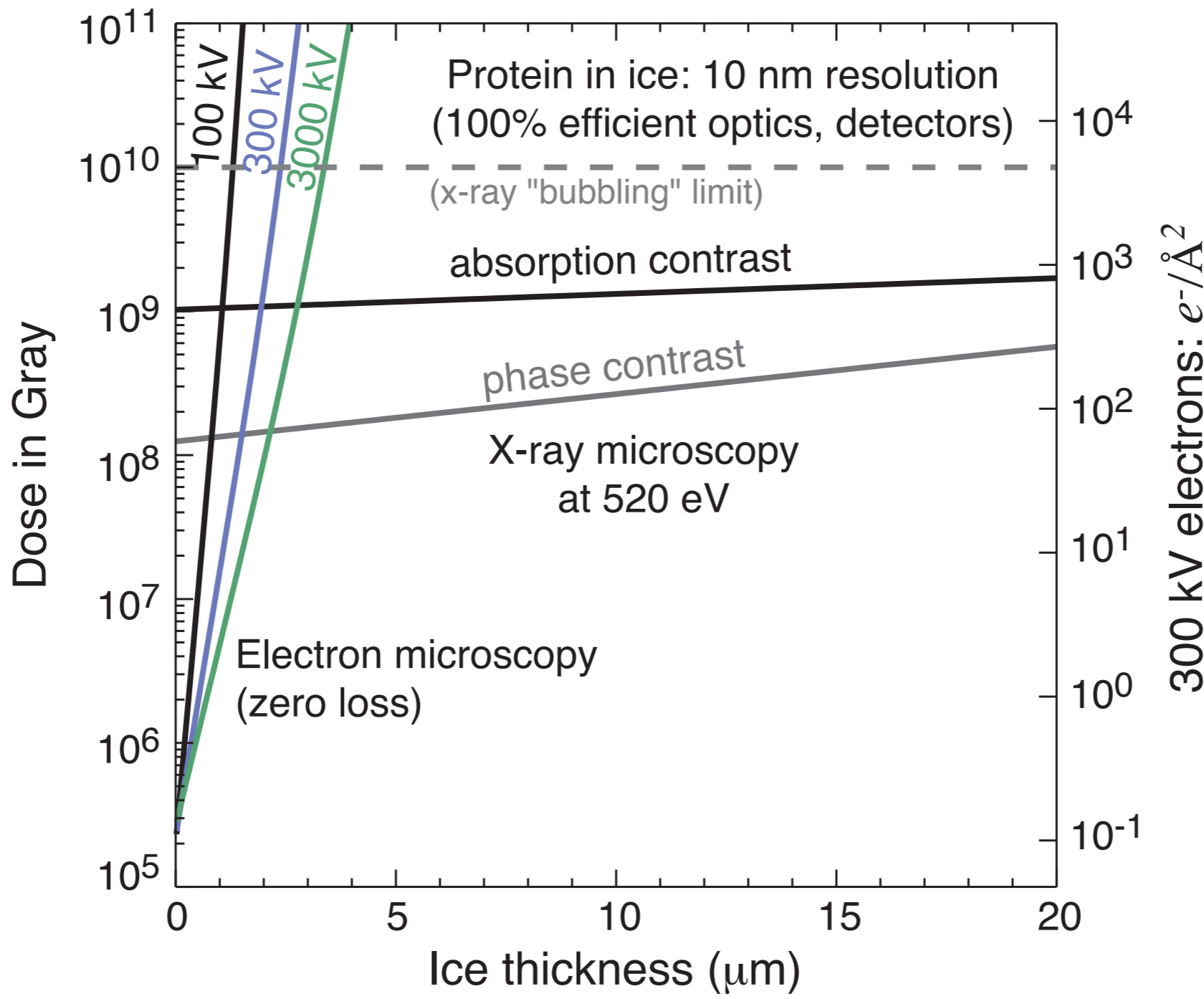


X. Huang *et al.*, *Optics Express* **17**, 13541 (2009)



# X rays are better than electrons for thick bio specimens

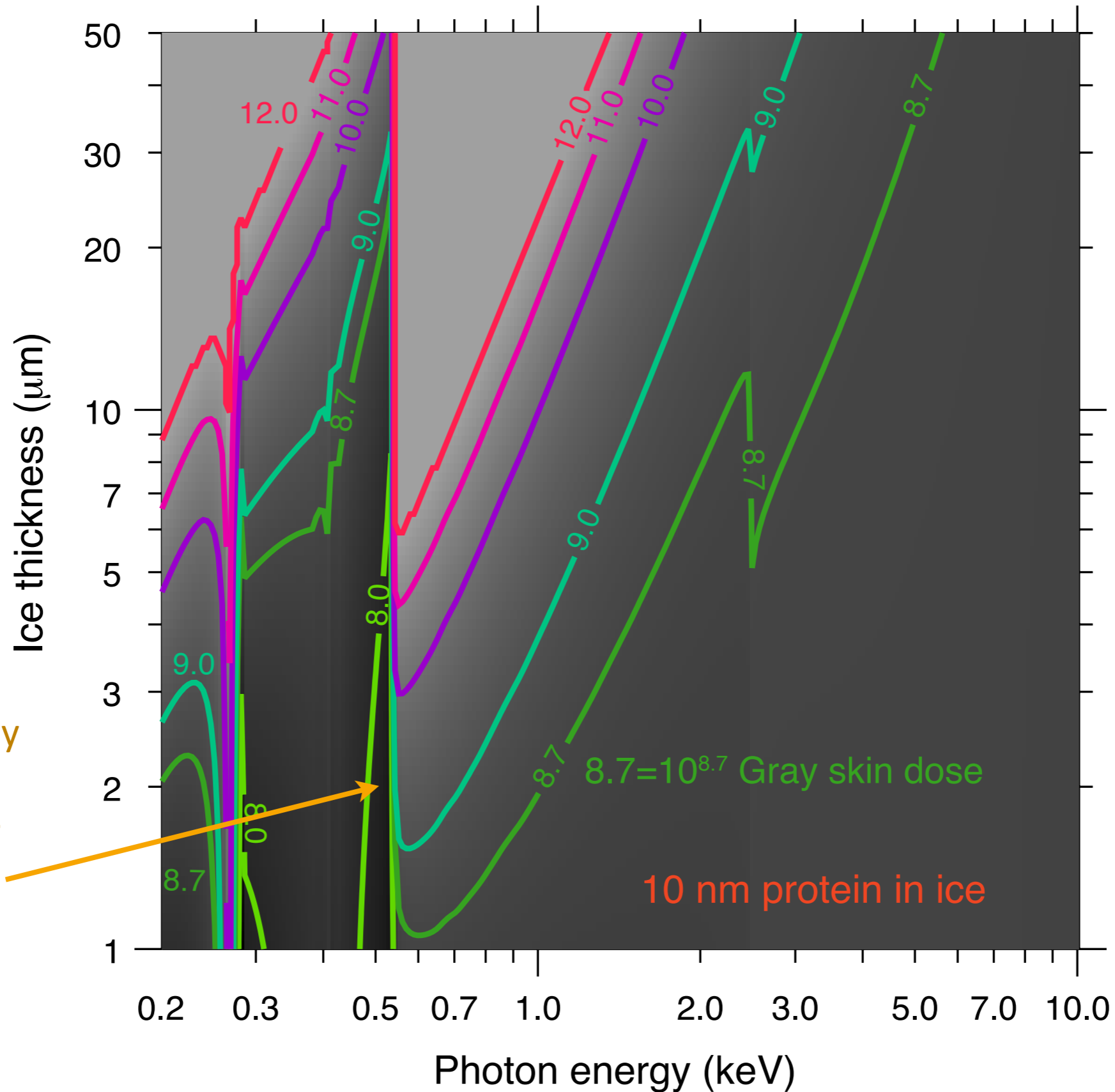
- Electrons are better for <500 nm thick specimens
- X rays are better for whole cells



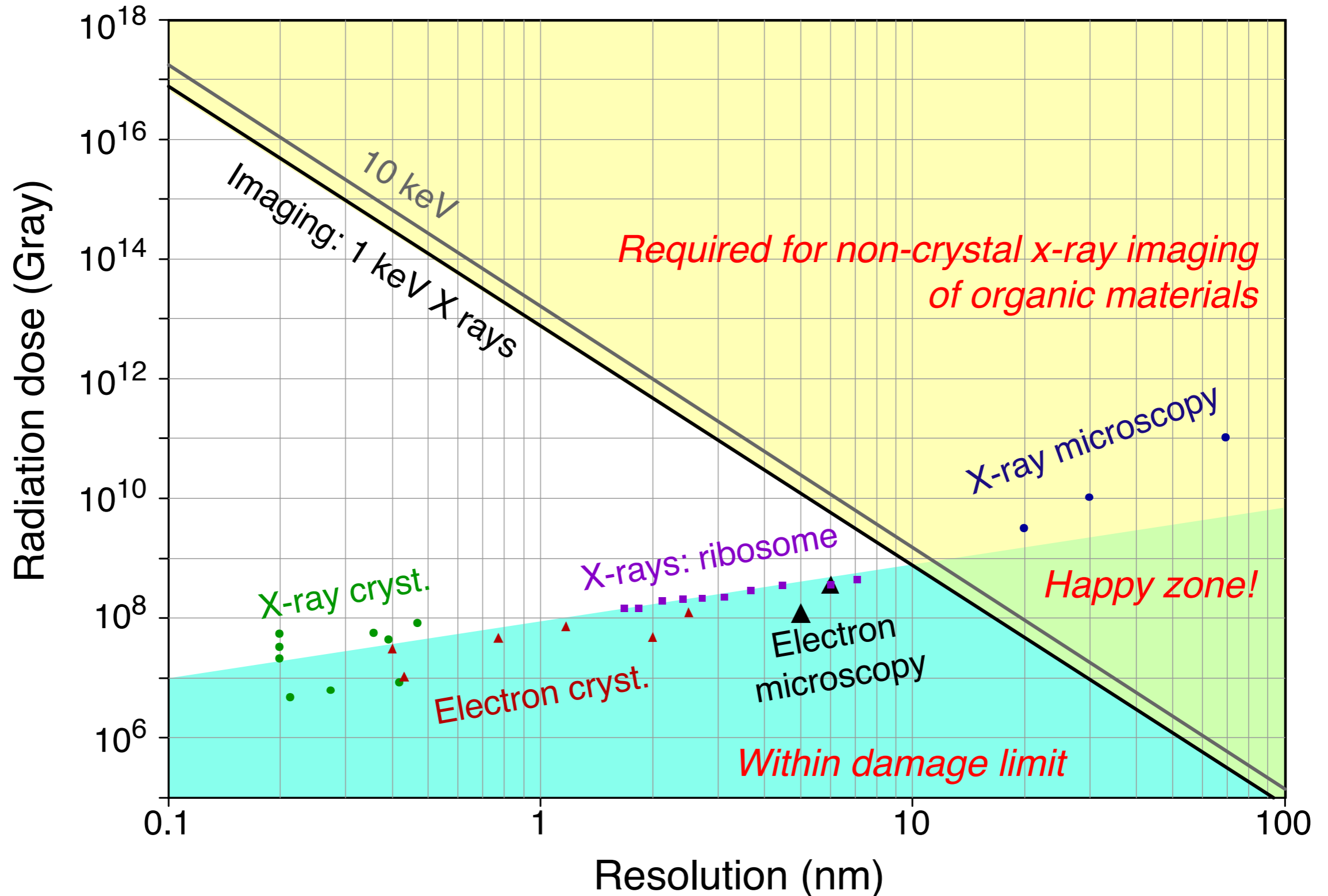
This plot: based on Jacobsen, Medenwaldt, and Williams, in **X-ray Microscopy & Spectromicroscopy** (Springer, 1998). See also Sayre *et al.*, *Science* **196**, 1339 (1977).



# Another look at dose versus energy and ice thickness



# What's the limit for cells?



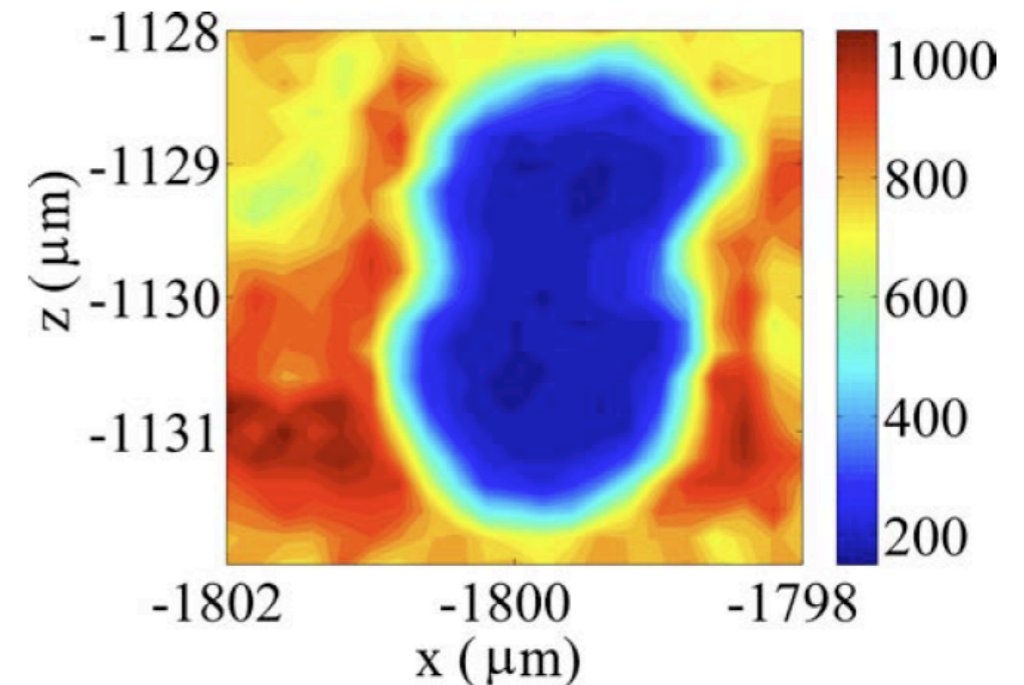
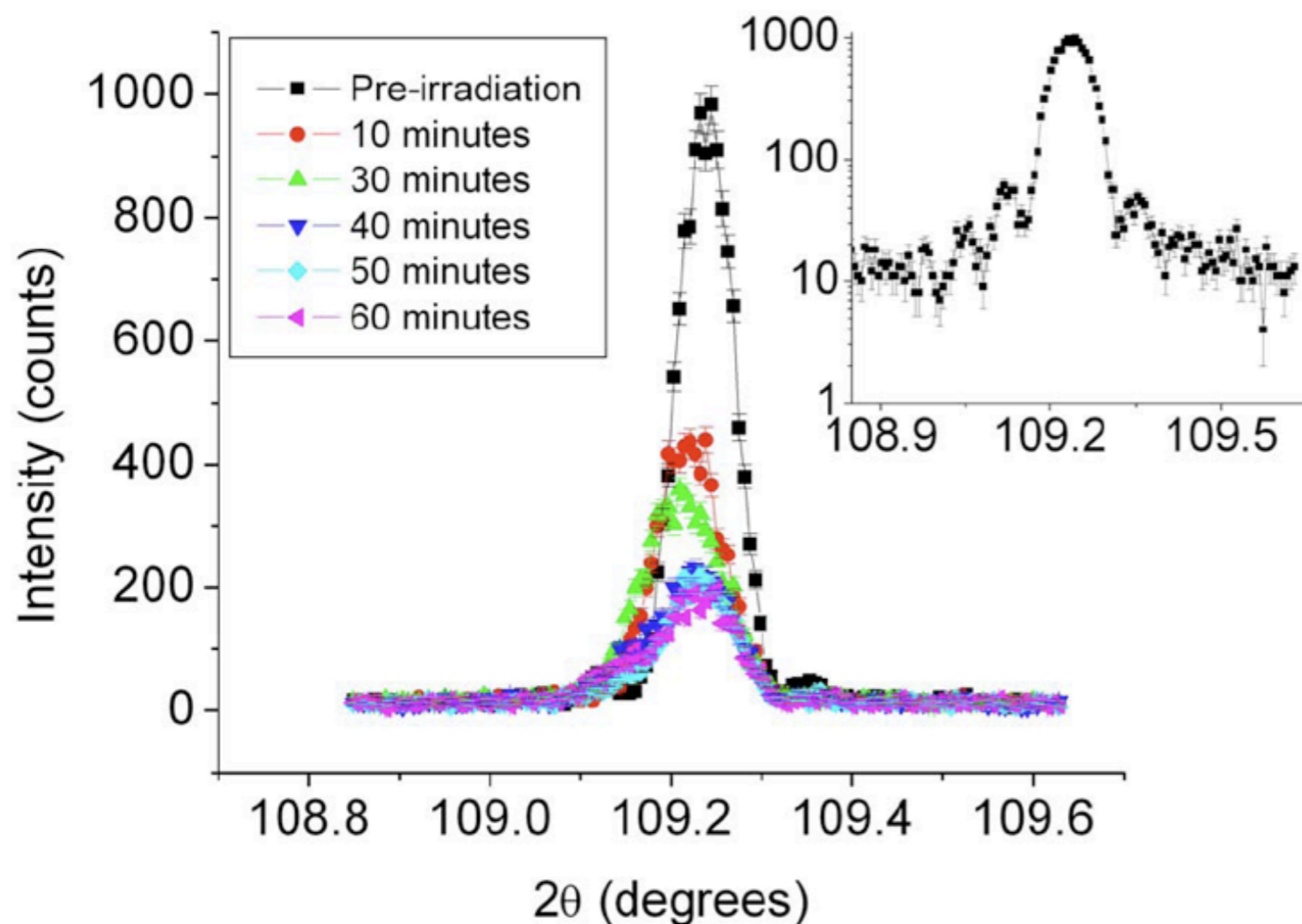
Howells et al., *JESRP* 170, 4 (2009)

See also Shen et al., *J. Sync. Rad.* **11**, 432 (2004)



# X-ray damage: Silicon in Silicon-on-insulator (SOI)

Non-recoverable fading of Si 008 diffraction peak in 140 nm thick SOI layer, using 11.2 keV X rays



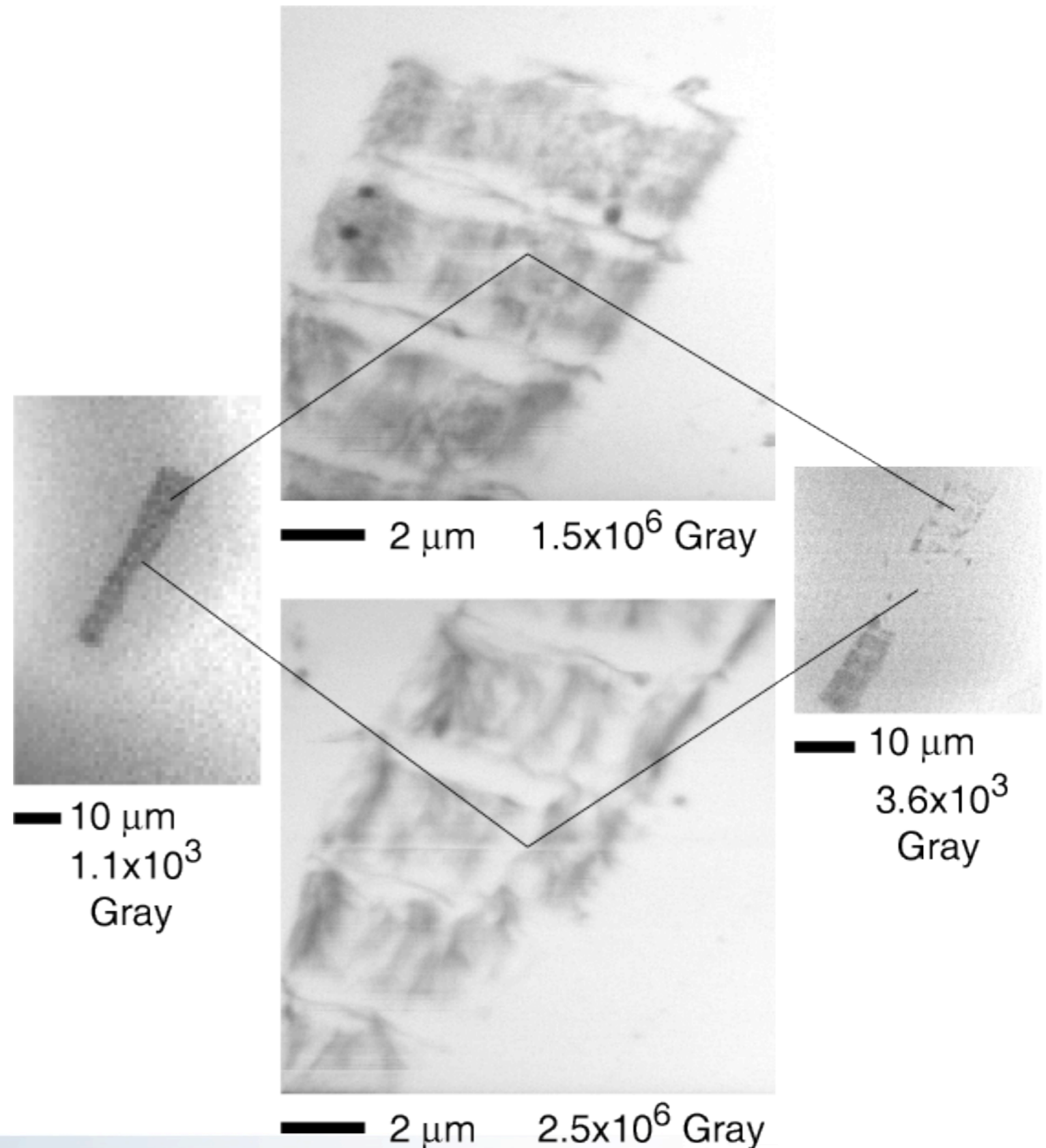
Beam spot:  $\sim 0.3 \mu\text{m}$   
Damage radius:  $\sim 1.8 \mu\text{m}$

$1.9 \times 10^8$  photons/sec into  $0.25 \times 0.30 \mu\text{m}$ , or  
 $\sim 6 \times 10^9$  Gray per 10 minutes

Polvino, Murray, Kalenci, Noyan, Lai, and Cai, *Appl. Phys. Lett.* **92**, 224105 (2008)

# Muscle damage

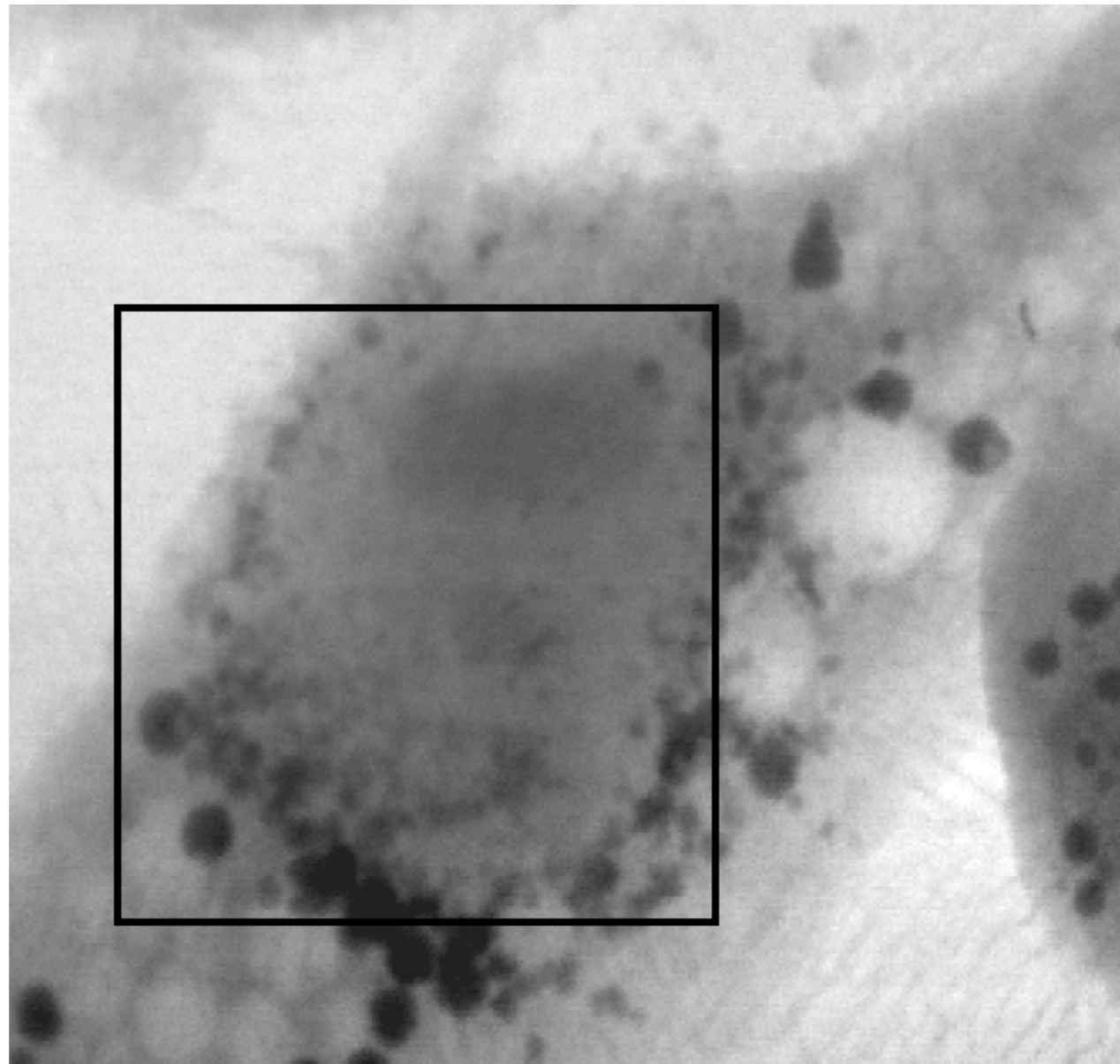
- Images: dragonfly flight muscle, with Clara Franzini-Armstrong
- At  $10^4$  Gray, myofibrils stop contracting in the presence of ATP. Bennett *et al.*, *J. Micros.* 172, 109 (1993)



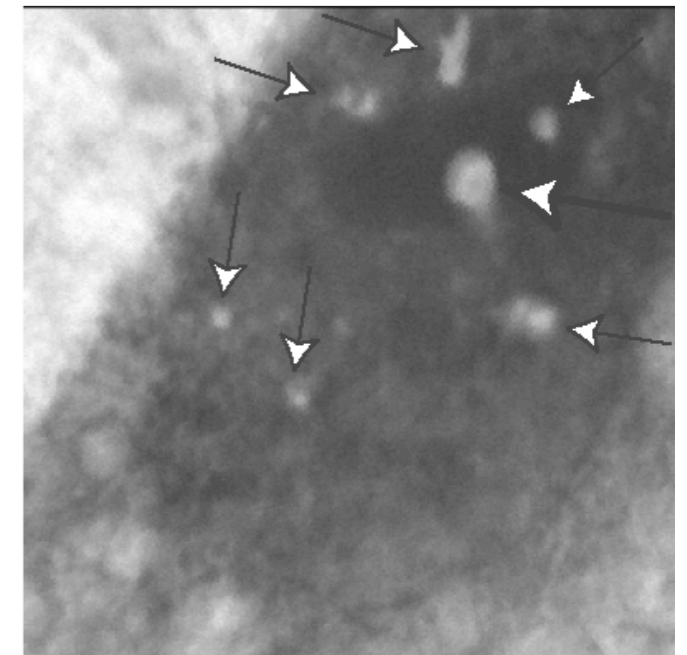
# Radiation damage resistance in cryo

Left: frozen hydrated image **after** exposing several regions to  $\sim 10^{10}$  Gray

Right: after warmup in microscope (eventually freeze-dried): holes indicate irradiated regions!



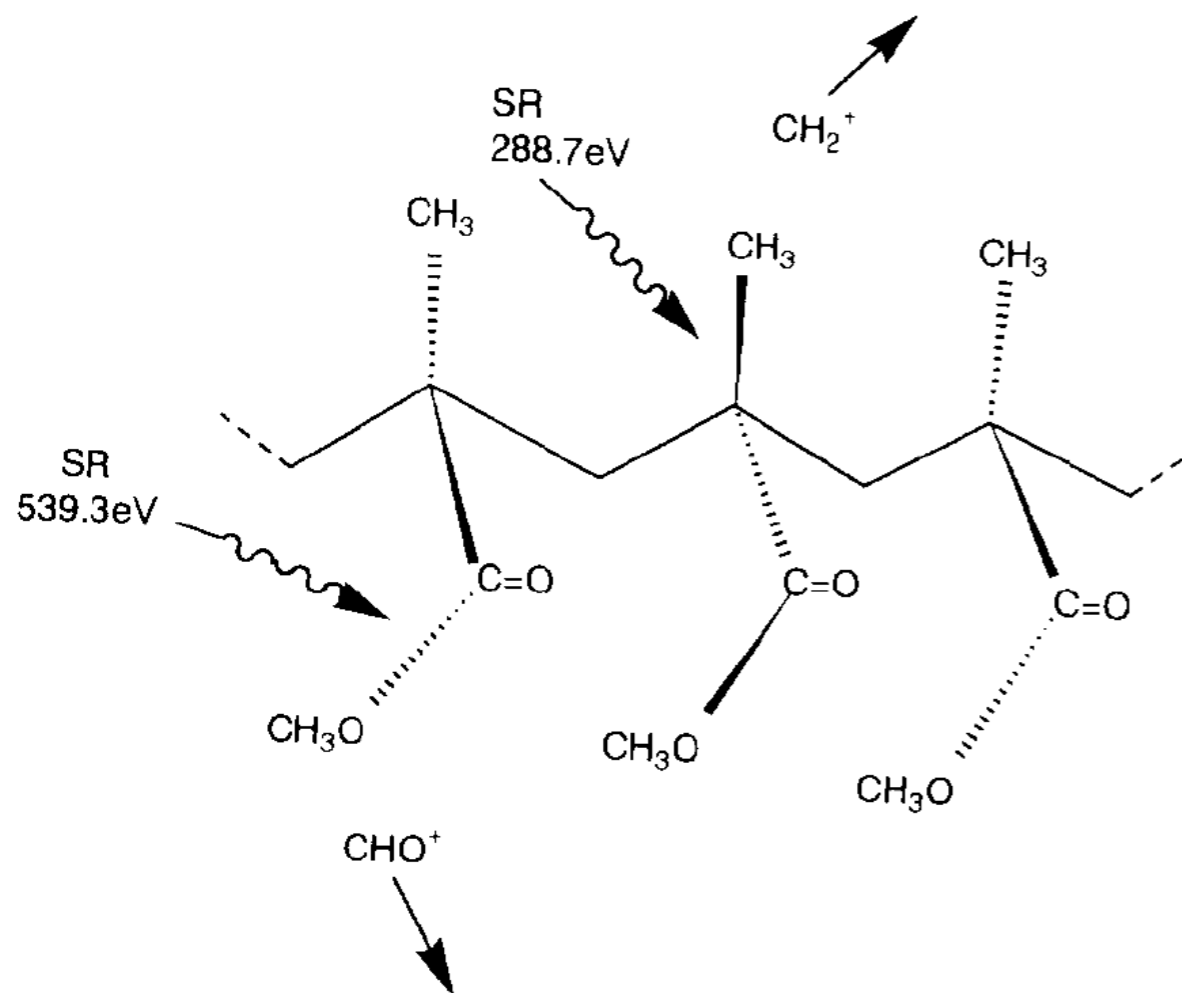
Maser *et al.*, *J. Microsc.* **197**, 68 (2000)



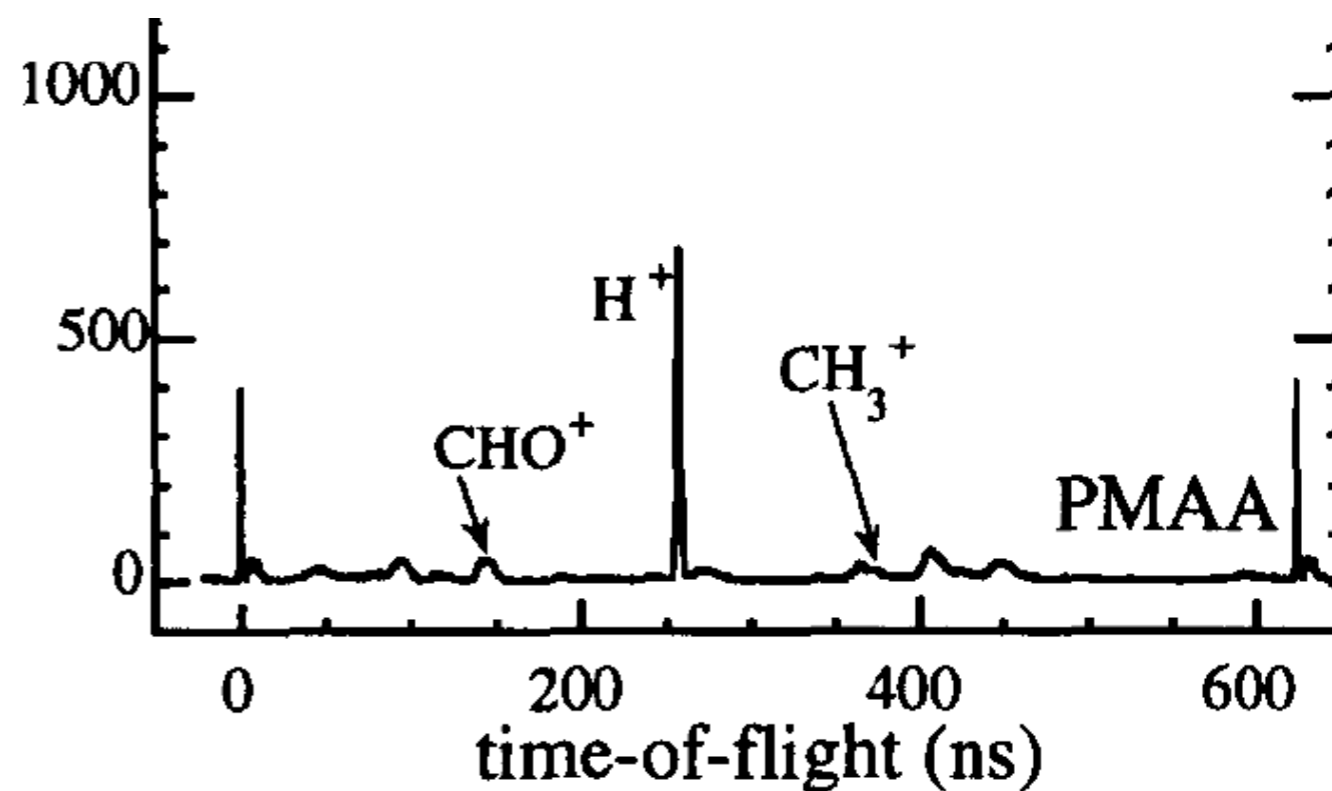
— 7  $\mu\text{m}$



# Radiation damage studies: poly (methyl methacrylate) or PMMA



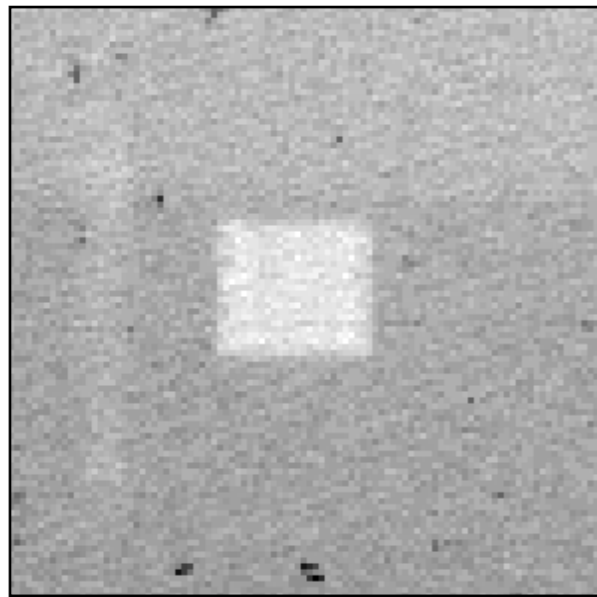
Tinone *et al.*, *Appl. Surf. Sci.* **79**, 89 (1994)



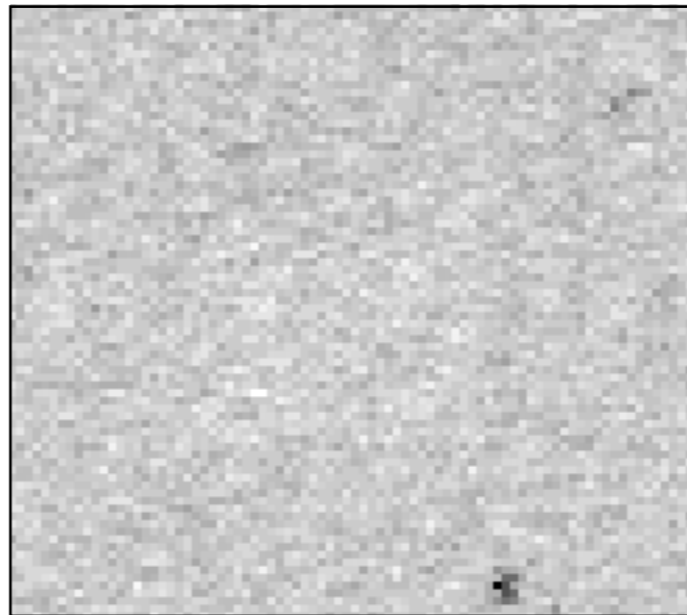
Tinone *et al.*, *J. Electron Spect. Rel. Phen.* **80**, 117 (1996).

# PMMA at room, LN2 temperature

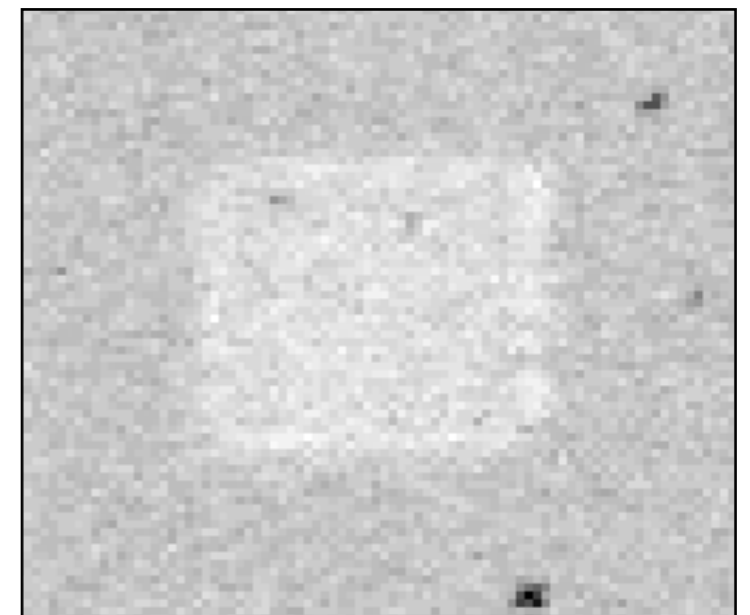
- Beetz and Jacobsen, J. Synchrotron Radiation **10**, 280 (2003)
- Repeated sequence: dose (small step size, long dwell time), spectrum (defocused beam)
- Images: dose region (small square) at end of sequence



Room temperature:  
mass loss  
immediately visible



LN2 temperature: no  
mass loss  
immediately visible

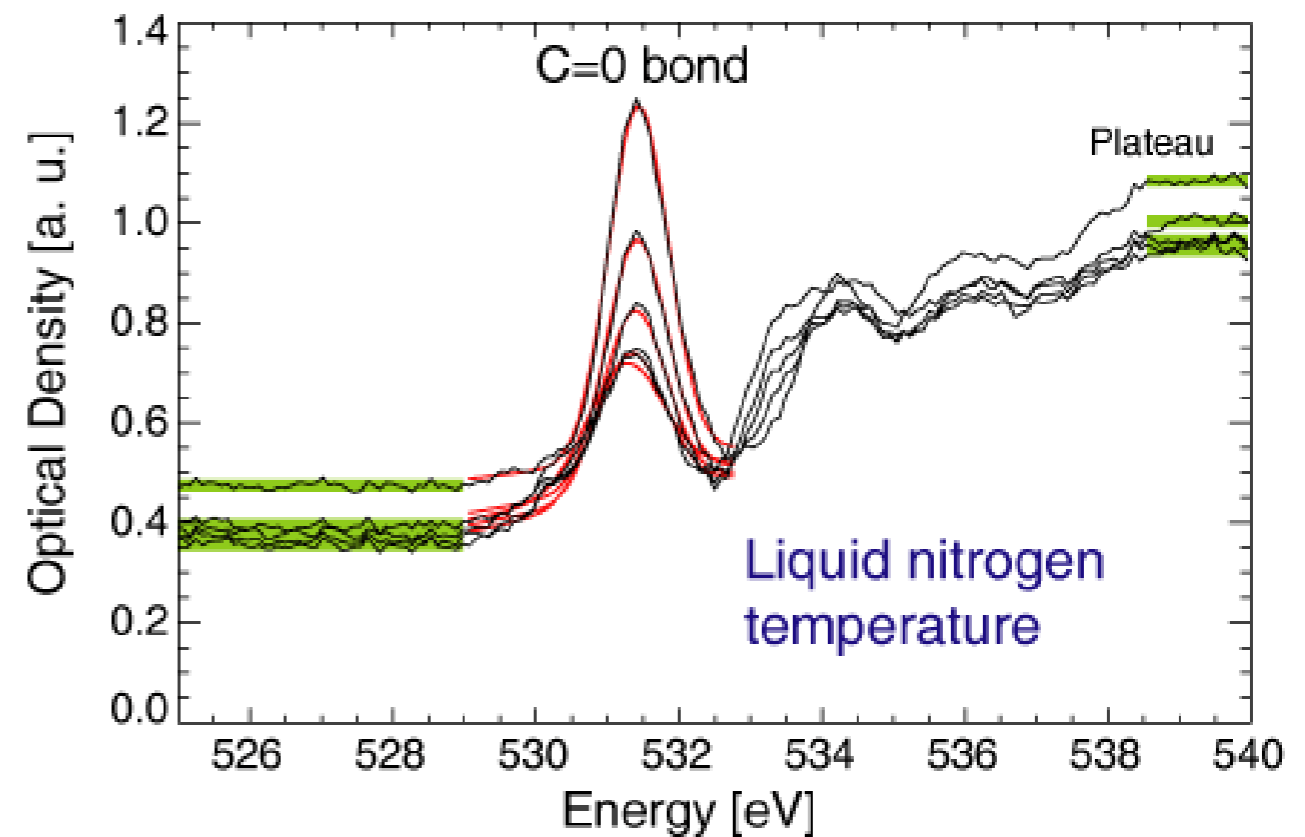
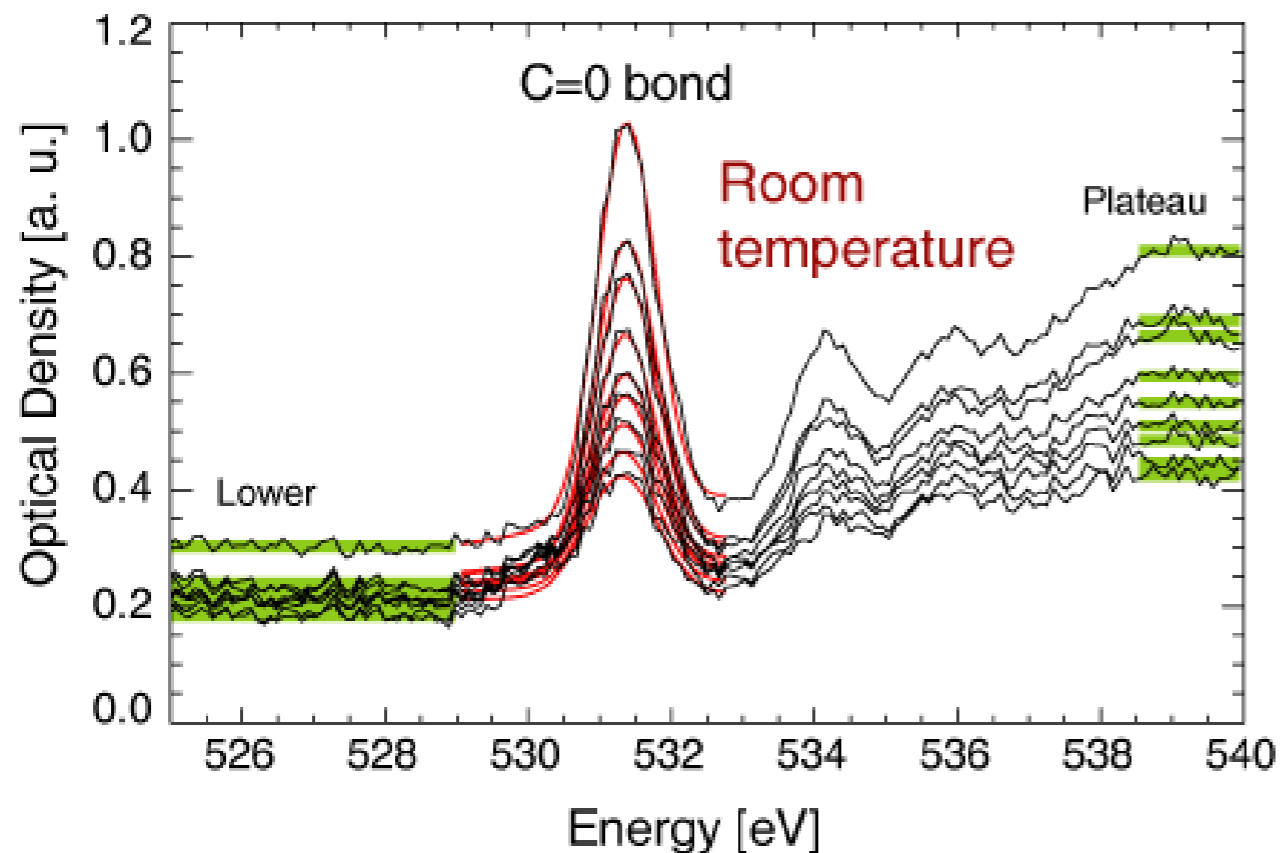


After warm-up: mass  
loss becomes visible



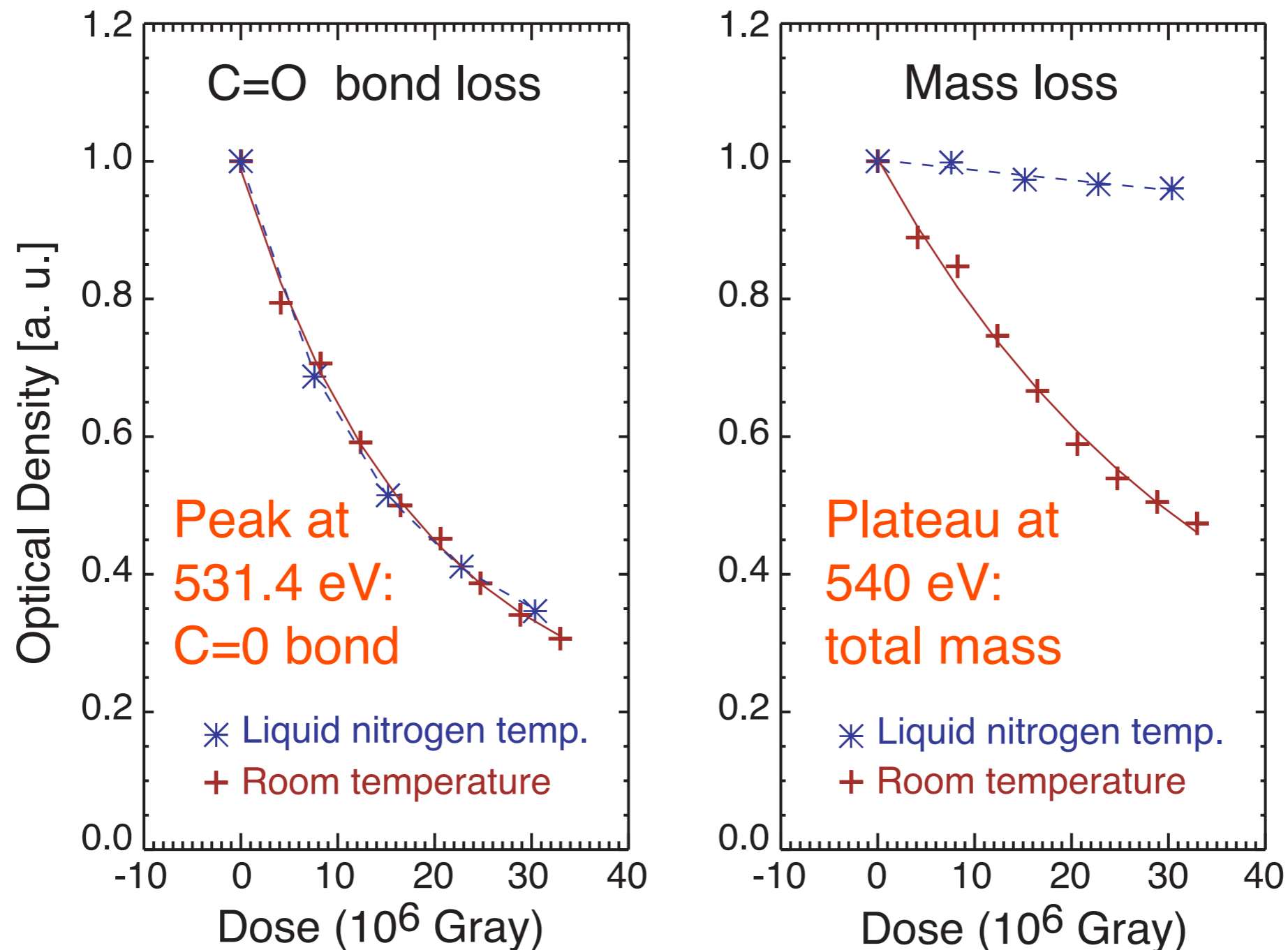
# PMMA at LN2, room temperature: XANES spectra

- Peak at 531.4 eV: C=O bond
- Plateau at 540 eV: total mass (plus some emphasis on oxygen  $\sigma^*$  bonds)
- Beetz and Jacobsen, *J. Synchrotron Radiation* **10**, 280 (2003)



# Cryo does not work miracles

LN<sub>2</sub> temp: protection against mass loss, but not against breaking bonds  
(at least C=O bond in dry PMMA)



Beetz and Jacobsen, *J. Synchrotron Radiation* **10**, 280 (2003)

# The Ramen noodle model of radiation damage



Macromolecular chains with no “encapsulating” matrix  
(dry, room temperature wet)

# The Ramen noodle model of radiation damage



Macromolecular chains in an “encapsulating” matrix  
(frozen hydrated)

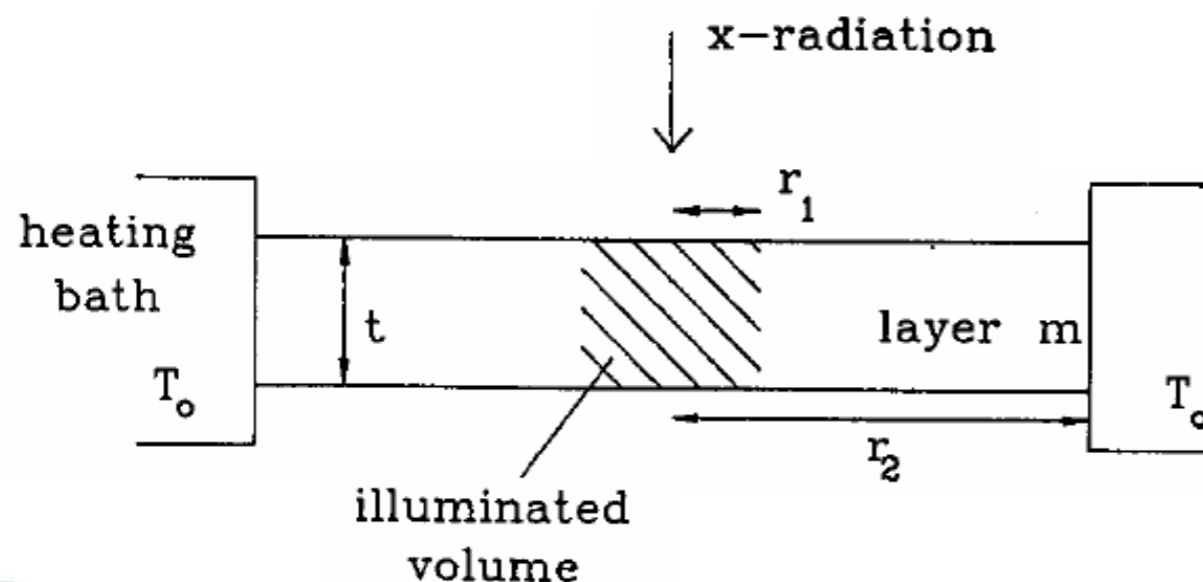
# The Ramen noodle model of radiation damage



Actual noodles *were* harmed during the filming of this movie

# Effects of $10^5$ photons in $(10 \text{ nm})^3$

- With no cooling, the temperature rises due to absorption:
  - $\text{H}_2\text{O}@500 \text{ eV} \Rightarrow 2300\text{K}$
  - $\text{H}_2\text{O}@3 \text{ keV} \Rightarrow 2200\text{K}$
  - $\text{Si}@10 \text{ keV} \Rightarrow 7300\text{K}$
- In scanning microscopes, localized heating with a thermal reservoir. Photon flux for  $\Delta T=1\text{K}$  in  $10 \text{ nm}$  wide spot with  $r_2=100 \mu\text{m}$ :
  - $\text{H}_2\text{O}@500 \text{ eV}: 4 \times 10^{10} \text{ photons/sec}$
  - $\text{Si}@10 \text{ keV}: 2 \times 10^{12} \text{ photons/sec}$



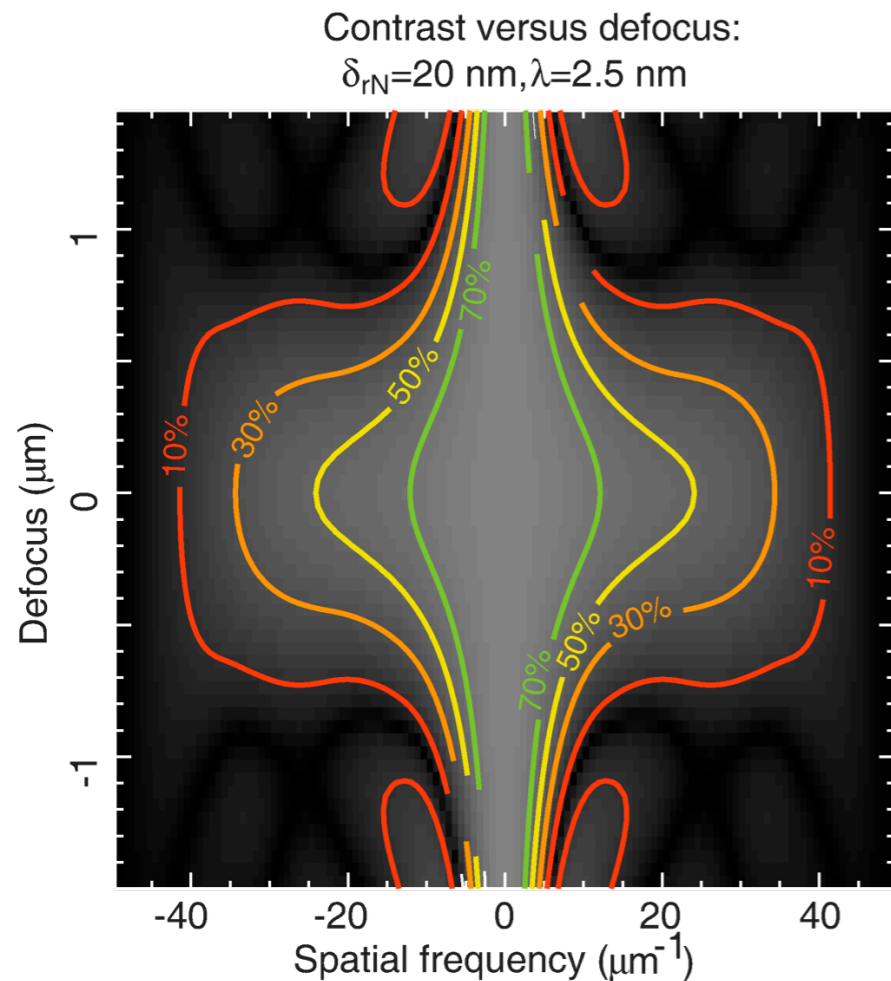
$$\Delta T = \frac{N}{t} \frac{h\nu \cdot \mu}{4\pi k} \left( 1 + 2 \ln \frac{r_2}{r_1} \right)$$

Greinke and Götz, XRM 1991

# Depth of focus in terms of zone plate/MLL $\Delta_{rN}$

Transverse:  $\Delta_t \Rightarrow \frac{\lambda}{4\theta} = \frac{\Delta_{rN}}{2}$

Longitudinal:  $\Delta_\ell \Rightarrow \frac{\lambda}{\theta^2} = 4\Delta_{rN} \frac{\Delta_{rN}}{\lambda}$



Incoherent brightfield imaging;  
 50% central stop.  
 Partial coherence: better?

